

IVIS Spectrum at DLAR Imaging Core

DLAR Imaging Core houses the [IVIS® Spectrum](#) advanced preclinical optical imaging system that enables non-invasive monitoring of cell trafficking, tumor growth and gene expression that enables generating visual and quantitative data for publications and grant submissions.

It enables:

- ✓ Non-invasive longitudinal studies
- ✓ High sensitivity for small signals
- ✓ Relatively quick imaging

Image capture

Back thinned, back illuminated grade 1 CCD Camera and Lens provides high quantum efficiency scanning over the entire visible to near-infrared spectrum. The system features 10 narrow band excitation filters: 415 nm – 760 nm and 18 narrow band emission filters: 490 nm – 850 nm. Lens provides high resolution - down to 20 microns at FOV of 4 cm.

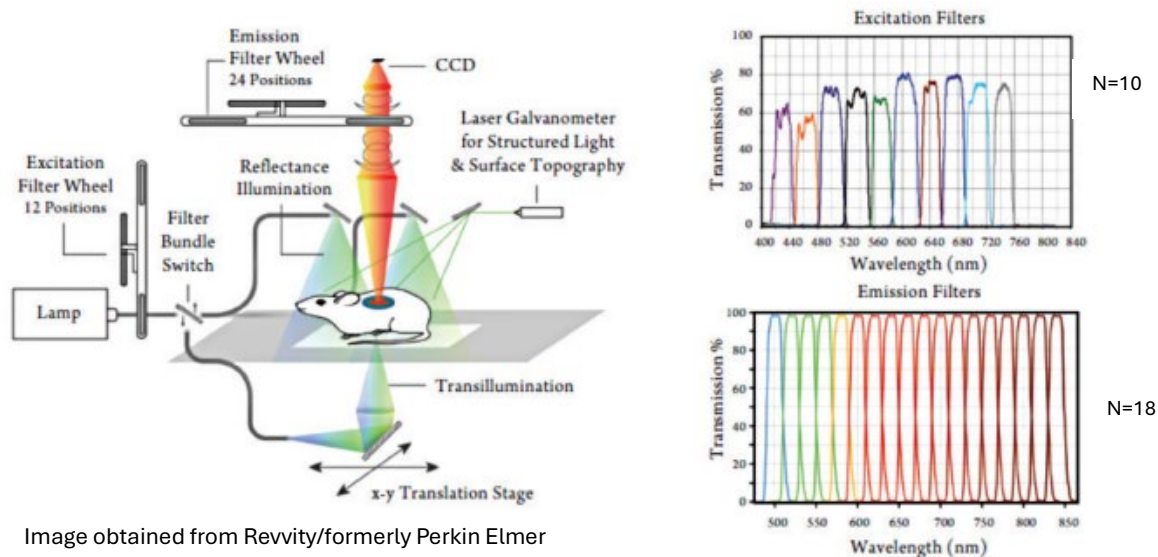
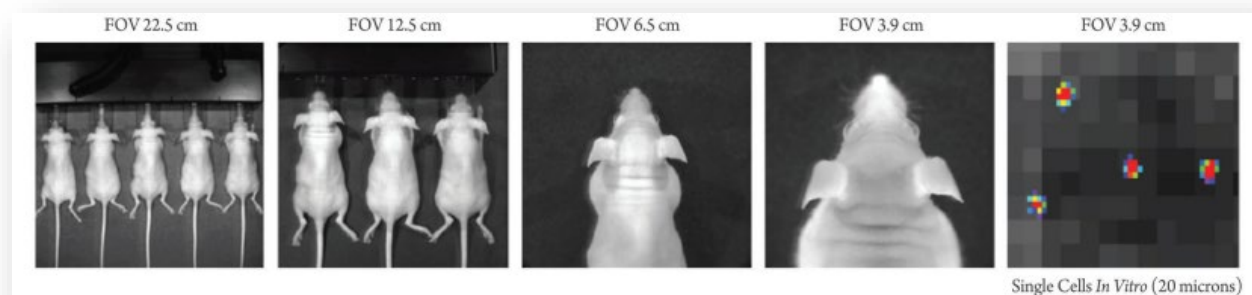


Image obtained from Revvity/formerly Perkin Elmer



From 20 microns to localize single cells to five whole mice, the IVIS Spectrum gives you the automated flexibility, throughput and resolution required to quantitate functional developments in whole animals down to a single cell. Image obtained from Revvity/formerly Perkin Elmer

Bioluminescence imaging- best in class in vivo sensitivity

Image multiple bioluminescent reporters like firefly luciferase, Renilla luciferase and bacterial luciferase *in vivo* at depth rapidly and quantitatively. The ultra-sensitive camera optics allows the detection of as few as five cells. [Revvity](#) offers Luciferin Luciferase Cell Lines Lentivirus Particles.

Fluorescence imaging - versatility in fluorescence

The IVIS Spectrum can image and quantify all commonly used fluorophores, including fluorescent proteins, dyes and conjugates. [Revvity](#) offers the broadest portfolio of fluorescent agents and dyes for *in vivo* applications. The IVIS Spectrum is the most sensitive system to visualize these fluorescent agents in various *in vivo* applications.

Multispectral Imaging with Advanced Spectral Unmixing Algorithms

Advanced spectral-unmixing algorithms and a broad range of high spectral resolution filter sets minimize autofluorescence and provide the opportunity to image a wide variety of targeted and activatable fluorescent probes and reporters.

Absolute localization in optical imaging - 3D analysis

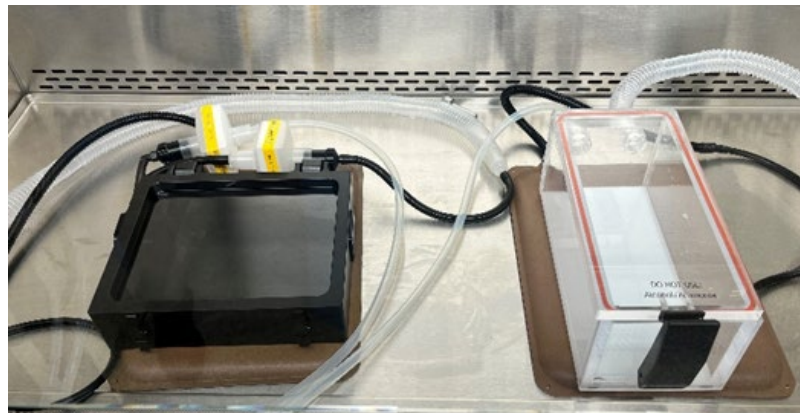
3D diffuse tomography utilizes structured light data with bioluminescence or fluorescence images to reconstruct three dimensional representations of light emitting reporters and compute signal strength.

Determine geometry and quantify the depth and intensity and of fluorescent sources in 3D space using FLIT (Fluorescent Imaging Tomography) or bioluminescent sources using DLIT (Diffused Luminescent Imaging Tomography).

XIC-3 Animal Isolation Chamber Kit

The chamber is available at DLAR, provides biological isolation of anesthetized mice or small rats before they are imaged in an *in vivo* imaging system.

It is equipped with HEPA filters allowing imaging of immunodeficient mice and mice infected with ABSL-2 level agents.



Application of IVIS spectrum Imaging System include:

- √ **Cancer biology:** Monitor tumor growth and metastasis using luciferase-tagged cells

- √ **Gene expression studies:** Track reporter gene expression (e.g., luciferase, GFP)
- √ **Infection models:** Visualize progression and clearance of bacterial or viral infections
- √ **Stem cell tracking:** Follow migration and survival of labeled cells post-transplant
- √ **Assessing therapeutic efficacy:** Evaluate drug delivery, targeting, and therapeutic effects
- √ **Immunology:** Study immune cell trafficking and inflammation

Image analysis

Living Image® is Revvity's proprietary software for in vivo optical imaging data acquisition and analysis, designed specifically to work with Revvity's IVIS family of imaging systems (e.g., IVIS Spectrum, IVIS Lumina). It enables researchers to visualize, quantify, and interpret bioluminescent and fluorescent imaging data collected. Offers tools to define regions of interest (ROIs) and extract quantitative signal data.

Living Image (LI) Acquisition/Analysis Package software allows for advanced quantification and analysis of data. Complimentary Living Image licenses are typically provided with IVIS systems upon request, for no additional cost.

List of publications and preprints that have used the IVIS Spectrum imaging system at the DLARIC

- Khan, M. I., Sankaran, K. R., & Rahaman, S. O. (2026). *Endosome-escaping engineered LNP-miR146a with in vivo biodistribution to mitigate inflammation and foreign body giant cell formation*. bioRxiv. <https://doi.org/10.64898/2026.02.13.705811>
- Bridgeman, C. J., Shen, R., McIlvaine, R. A., Edwards, C., Ackun-Farmmer, M. A., & Jewell, C. M. (2024). Synthetic organic materials for targeting immunotherapies to lymph nodes. *Chemistry of Materials*, 36(19), 9031–9045. <https://doi.org/10.1021/acs.chemmater.4c00947>
- Edwards, C., Carey, S. T., & Jewell, C. M. (2023). Harnessing biomaterials to study and direct antigen-specific immunotherapy. *ACS Applied Bio Materials*, 6(6), 2017–2028. <https://doi.org/10.1021/acsabm.3c00136>
- Andorko, J. I., Tsai, S. J., Gammon, J. M., Carey, S. T., Zeng, X., Gosselin, E. A., Edwards, C., Shah, S. A., Hess, K. L., & Jewell, C. M. (2022). Spatial delivery of immune cues to lymph nodes to define therapeutic outcomes in cancer vaccination. *Biomaterials Science*, 10(17), 4612–4626. <https://doi.org/10.1039/D2BM00403H>
- Edwards, C., Shah, S. A., Gebhardt, T., & Jewell, C. M. (2023). Exploiting Unique Features of Microneedles to Modulate Immunity. *Advanced materials (Deerfield Beach, Fla.)*, 35(52), e2302410. <https://doi.org/10.1002/adma.202302410>
- Rui, Y., Eppler, H. B., Yanes, A. A., & Jewell, C. M. (2023). Tissue-Targeted Drug Delivery Strategies to Promote Antigen-Specific Immune Tolerance. *Advanced healthcare materials*, 12(6), e2202238. <https://doi.org/10.1002/adhm.202202238>
- Ackun-Farmmer, M. A., & Jewell, C. M. (2023). Delivery route considerations for designing antigen-specific biomaterial strategies to combat autoimmunity. *Advanced nanobiomed research*, 3(3), 2200135. <https://doi.org/10.1002/anbr.202200135>

- Gammon, J.M., Carey, S.T., Saxena, V. *et al.* Engineering the lymph node environment promotes antigen-specific efficacy in type 1 diabetes and islet transplantation. *Nat Commun* **14**, 681 (2023).
<https://doi.org/10.1038/s41467-023-36225-5>
- Carey, S. T., Bridgeman, C., & Jewell, C. M. (2023). Biomaterial strategies for selective immune tolerance: Advances and gaps. *Advanced Science*, *10*, 2205105. <https://doi.org/10.1002/advs.202205105>
- Ackun-Farmmer, M. A., Willson Shirkey, M., Oakes, R. S., Shah, S. A., Edwards, C., Kapnick, S., Carey, S. T., Yanes, A., Bromberg, J., & Jewell, C. M. (2024). Engineered Immune Constructs Alter Antigen-Specific Immune Tolerance and Confer Durable Protection in Myelin-Driven Autoimmunity. *ACS nano*, *18*(46), 31780–31793. <https://doi.org/10.1021/acsnano.4c06667>
- Tsai, S. J., Kapnick, S. M., Carey, S. T., McIlvaine, R. A., Rui, Y., Eppler, H. B., Shah, S. A., Bridgeman, C. J., Yanes, A. A., Black, S. K., Zeng, X., Gammon, J. M., & Jewell, C. M. (2026). Defined distribution and features of lymph node therapies enable recruitment and manipulation of antigen-specific T cell response. *Molecular therapy : the journal of the American Society of Gene Therapy*, *34*(2), 1009–1023. <https://doi.org/10.1016/j.ymthe.2025.10.026>
- McIlvaine, R. A., Kapnick, S. M., Carey, S. T., & Jewell, C. M. (2025). Regulation of response to antigen peptides is independent of peptide distribution in lymph node therapeutics. *Biomaterials science*, *13*(19), 5538–5549. <https://doi.org/10.1039/d5bm00328h>
- McCright, J., Skeen, C., Yarmovsky, J., & Maisel, K. (2022). Nanoparticles with dense poly(ethylene glycol) coatings with near neutral charge are maximally transported across lymphatics and to the lymph nodes. *Acta biomaterialia*, *145*, 146–158. <https://doi.org/10.1016/j.actbio.2022.03.054>
- Yeruva, T., Morris Iii, R. J., Kumar, S., Zhao, L., Kofinas, P., & Duncan, G. A. (2025). Rapid *in situ* forming PEG hydrogels for mucosal drug delivery. *Biomaterials science*, *13*(16), 4390–4399. <https://doi.org/10.1039/d4bm01101e>
- Yeruva, T., Yang, S., Kaluzienski, M., Louisthelmy, R., Doski, S., Maisel, K., & Duncan, G. A. (2025). Synthetic Mucus Biomaterials Enable Localized Therapeutic Antibody Delivery in Inflammatory Bowel Disease. *bioRxiv : the preprint server for biology*, 2025.08.15.670558. <https://doi.org/10.1101/2025.08.15.670558>
- Ou, W., Stewart, S., White, A., *et al.* (2023). In-situ cryo-immune engineering of tumor microenvironment with cold-responsive nanotechnology for cancer immunotherapy. *Nature Communications*, *14*, Article 392. <https://doi.org/10.1038/s41467-023-36045-7>
- Xu, J., Liu, Y., Liu, S., Ou, W., White, A., Stewart, S., Tkaczuk, K. H. R., Ellis, L. M., Wan, J., Lu, X., & He, X. (2022). Metformin Bicarbonate-Mediated Efficient RNAi for Precise Targeting of *TP53* Deficiency in Colon and Rectal Cancers. *Nano today*, *43*, 101406. <https://doi.org/10.1016/j.nantod.2022.101406>
- Quinlan, J. A., Inglut, C. T., Srivastava, P., Rahman, I., Stabile, J., Gaitan, B., Arnau Del Valle, C., Baumiller, K., Gaur, A., Chiou, W. A., Karim, B., Connolly, N., Robey, R. W., Woodworth, G. F., Gottesman, M. M., & Huang, H. C. (2024). Carrier-Free, Amorphous Verteporfin Nanodrug for Enhanced Photodynamic Cancer Therapy and Brain Drug Delivery. *Advanced science (Weinheim, Baden-Wurttemberg, Germany)*, *11*(17), e2302872. <https://doi.org/10.1002/advs.202302872>
- Yang, J., Ma, C. H., Quinlan, J. A., McNaughton, K., Lee, T., Shin, P., Hauser, T., Kaluzienski, M. L., Vig, S., Quang, T. T., Starost, M. F., Huang, H. C., & Mueller, J. L. (2024). Light-activatable minimally invasive ethyl cellulose ethanol ablation: Biodistribution and potential applications. *Bioengineering & translational medicine*, *9*(6), e10696. <https://doi.org/10.1002/btm2.10696>
- Ma, C. H., Yang, J., Quinlan, J. A., McNaughton, K., Kaluzienski, M. L., Hauser, T., Starost, M. F., Mueller, J. L., & Huang, H. C. (2025). Synergizing photodynamic therapy and ethanol ablation: Light-activatable sustained-exposure ethanol injection technology for enhanced tumor ablation. *Bioengineering & translational medicine*, *10*(5), e70028. <https://doi.org/10.1002/btm2.70028>
- Goldstein, M. E., Ignacio, M. A., Loube, J. M., Whorton, M. R., & Scull, M. A. (2024). Human Stimulator of Interferon Genes Promotes Rhinovirus C Replication in Mouse Cells In Vitro and In Vivo. *Viruses*, *16*(8), 1282. <https://doi.org/10.3390/v16081282>